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APPLICATION NO.	FILING DATE	FIRST NAMED INVENTOR	ATTORNEY DOCKET NO.	CONFIRMATION NO.
10/812,268	03/29/2004	Jeffrey William Mochlenbruck	2103.013882/SB1064US3DIV	2977
45488	7590	03/28/2008		
WILLIAMS, MORGAN & AMERSON 10333 RICHMOND, SUITE 1100 HOUSTON, TX 77042			EXAMINER TSAY, MARSHA M	
			ART UNIT	PAPER NUMBER
			1656	
			MAIL DATE	DELIVERY MODE
			03/28/2008	PAPER

Please find below and/or attached an Office communication concerning this application or proceeding.

The time period for reply, if any, is set in the attached communication.

Office Action Summary

Application No.

10/812,268

Applicant(s)

MOEHLENBRUCK ET AL.

Examiner

Marsha M. Tsay

Art Unit

1656

-- The MAILING DATE of this communication appears on the cover sheet with the correspondence address --
Period for Reply

A SHORTENED STATUTORY PERIOD FOR REPLY IS SET TO EXPIRE 3 MONTH(S) OR THIRTY (30) DAYS, WHICHEVER IS LONGER, FROM THE MAILING DATE OF THIS COMMUNICATION.

- Extensions of time may be available under the provisions of 37 CFR 1.136(a). In no event, however, may a reply be timely filed after SIX (6) MONTHS from the mailing date of this communication.
- If NO period for reply is specified above, the maximum statutory period will apply and will expire SIX (6) MONTHS from the mailing date of this communication.
- Failure to reply within the set or extended period for reply will, by statute, cause the application to become ABANDONED (35 U.S.C. § 133). Any reply received by the Office later than three months after the mailing date of this communication, even if timely filed, may reduce any earned patent term adjustment. See 37 CFR 1.704(b).

Status

- 1) ☒ Responsive to communication(s) filed on 23 January 2008.
- 2a) ☒ This action is **FINAL**. 2b) ☐ This action is non-final.
- 3) ☐ Since this application is in condition for allowance except for formal matters, prosecution as to the merits is closed in accordance with the practice under *Ex parte Quayle*, 1935 C.D. 11, 453 O.G. 213.

Disposition of Claims

- 4) ☒ Claim(s) 82-102 and 125 is/are pending in the application.
- 4a) Of the above claim(s) _____ is/are withdrawn from consideration.
- 5) ☐ Claim(s) _____ is/are allowed.
- 6) ☒ Claim(s) 82-102 and 125 is/are rejected.
- 7) ☐ Claim(s) _____ is/are objected to.
- 8) ☐ Claim(s) _____ are subject to restriction and/or election requirement.

Application Papers

- 9) ☐ The specification is objected to by the Examiner.
- 10) ☐ The drawing(s) filed on _____ is/are: a) ☐ accepted or b) ☐ objected to by the Examiner.
Applicant may not request that any objection to the drawing(s) be held in abeyance. See 37 CFR 1.85(a).
Replacement drawing sheet(s) including the correction is required if the drawing(s) is objected to. See 37 CFR 1.121(d).
- 11) ☐ The oath or declaration is objected to by the Examiner. Note the attached Office Action or form PTO-152.

Priority under 35 U.S.C. § 119

- 12) ☐ Acknowledgment is made of a claim for foreign priority under 35 U.S.C. § 119(a)-(d) or (f).
- a) ☐ All b) ☐ Some * c) ☐ None of:
1. ☐ Certified copies of the priority documents have been received.
 2. ☐ Certified copies of the priority documents have been received in Application No. _____.
 3. ☐ Copies of the certified copies of the priority documents have been received in this National Stage application from the International Bureau (PCT Rule 17.2(a)).

* See the attached detailed Office action for a list of the certified copies not received.

Attachment(s)

- 1) ☐ Notice of References Cited (PTO-892)
- 2) ☐ Notice of Draftsperson's Patent Drawing Review (PTO-948)
- 3) ☒ Information Disclosure Statement(s) (PTO/S5108)
Paper No(s)/Mail Date 02.29.08
- 4) ☐ Interview Summary (PTO-413)
Paper No(s)/Mail Date _____
- 5) ☐ Notice of Informal Patent Application
- 6) ☐ Other: _____

A request for continued examination under 37 CFR 1.114, including the fee set forth in 37 CFR 1.17(e), was filed in this application after final rejection. Since this application is eligible for continued examination under 37 CFR 1.114, and the fee set forth in 37 CFR 1.17(e) has been timely paid, the finality of the previous Office action has been withdrawn pursuant to 37 CFR 1.114. Applicant's submission filed on January 23, 2008 has been entered.

Claims 1-81, 103-124 are canceled. Claims 82-102, 125 are pending and currently under examination.

Applicants' arguments have been fully considered and are deemed to be persuasive to overcome some of the rejections previously applied. Rejections and/or objections not reiterated from previous Office actions are hereby withdrawn.

Priority: The priority date is April 7, 2000.

Objections and Rejections

Claim 83 is objected to because of the following informalities: in claim 83, line 2, the term "xenogeneic" should be corrected to "xenogenic". Appropriate correction is required.

The following is a quotation of 35 U.S.C. 103(a) which forms the basis for all obviousness rejections set forth in this Office action:

(a) A patent may not be obtained though the invention is not identically disclosed or described as set forth in section 102 of this title, if the differences between the subject matter sought to be patented and the prior art are such that the subject matter as a whole would have been obvious at the time the invention was made to a person having ordinary skill in the art to which said subject matter pertains. Patentability shall not be negated by the manner in which the invention was made.

Claims 82-88, 91-102, 125 are rejected under 35 U.S.C. 103(a) as being unpatentable over Gan et al. (US 5964807; previously cited) in view of Mechanic (US 5854397; previously cited) as evidenced by Matsuzaki et al. (1996 Spine 21(2): 178-183; IDS 03/29/04).

Gan et al. disclose a method of manufacturing an implantable hybrid material comprising intervertebral disc cells and a biodegradable support substrate for placement into the intervertebral disc space (col. 5 lines 42-44). The intervertebral disc cells are nucleus pulposus cells extracted from the nucleus pulposus of lumbar discs, sacral discs, or cervical discs (col. 8 lines 1-7). Gan et al. further disclose the cells may be extracted from donor tissue (col. 8 lines 8-9) by surgical techniques (col. 8 lines 39-44). The extracted nucleus pulposus tissue can be treated with enzymes to disaggregate the cells (col. 8 lines 50-51). Further, the isolated intervertebral disc cells can be cultured alone or seeded onto a biodegradable substance and cultured together with the biodegradable substance for later implantation (col. 8 lines 58-61). To prepare the hybrid material, intervertebral disc cells are combined with biodegradable substrate materials, i.e. polymer foams (col. 7 lines 11-20, lines 65-66). Further, the hybrid material can also include factors to enhance cell growth, i.e. TGF- β , EGF (col. 8 lines 62-64). Gan et al. do not teach cross-linking.

Mechanic discloses a process for cross-linking a proteinaceous material, including collagen, collagen fibrils, and collagen matrices (col. 4 lines 15-16). According to Mechanic, the term proteinaceous material includes both proteins such as collagen and protein-containing materials such as tissue (col. 4 lines 19-20). Proteinaceous materials soaked in a first media solution and irradiated in a second are better cross-linked, show improved mechanical properties and decreased susceptibility to proteolytic degradation (col. 5 lines 1-4). Mechanic discloses

solutions of high osmolality are generally used for the first media solution, i.e. sodium, chloride, potassium buffers, and Good's buffers, where in the osmolality have been increased by addition of a solute, such as sucrose (col. 5 line 10). In working examples 1-10, Mechanic discloses proteinaceous materials from different sources to be crosslinked, including pericardium tissue, collagen fibrils, and collagen (col. 8-13). In example 8, rat type I collagen was divided into six samples and each sample was placed in a dialysis bag with 300 mg NaCl (col. 12 line 35-37). Samples 5-6 were dialyzed into phosphate buffered saline pH 7.4 including 50% sucrose, and 0.2% methylene blue (col. 12, lines 40-41) and then exposed to a white floodlight while holding the temperature between 8° and 12°C (col. 12, lines 45-50).

It is known in the art that living, intact nucleus pulposus cells actively synthesize collagen (p. 179) (Matsuzaki et al. 1996 Spine 21(2): 178-183; IDS 03/29/04).

It would have been obvious to one of ordinary skill in the art at the time the invention was made to modify the teachings of Gan et al. by adding the cross-linking step of Mechanic et al. to the nucleus pulposus tissue culture of Gan et al. for a method of manufacturing an implantable material comprising nucleus pulposus tissue and a biodegradable support substrate for placement into the intervertebral disc space (claims 82-88, 91-100). The motivation to do so is given by Mechanic, which teaches cross-linking tissue and/or collagen results in a stable, bio-product that resists in vivo degradation and calcification when implanted; therefore, one of ordinary skill would expect to be successful in manufacturing a more structurally stable disc implant by cross-linking the nucleus pulposus cells of Gan et al. since it is known in the art that said cells synthesize collagen.

It would also have been obvious to one of ordinary skill in the art at the time the invention was made to add an additional therapeutic substance, i.e. TGF- β , to the disc implant manufactured by the method of Gan et al. in view of Mechanic (claims 101-102, 125). The motivation to do so is given by Gan et al., which teach that additional therapeutic substances can be added to the nucleus pulposus hybrid implant and may enhance growth of intervertebral disc cells in the recipient.

In their remarks, Applicants assert Gan et al. teach the use of donor nucleus pulposus cells, but not tissue harvested from a donor *per se*. Although Gan notes "tissue may be extracted from the nucleus pulposus of lumbar discs, sacral discs and cervical discs," the collagen and other components of that tissue subsequently are discarded to obtain isolated nucleus pulposus cells. These isolated cells are then combined with materials intended to substitute for the discarded nucleus pulposus tissue collagen and other components from which they were isolated. Throughout the specification, Gan repeatedly emphasizes the mechanical and chemical destruction of the nucleus pulposus tissue to liberate cells for use in creating a hybrid matrix containing bioactive glass, polymer foam, and polymer foam coated with a sol gel bioactive material (col. 6, lines 30-32). Gan also makes no mention of crosslinking nucleus pulposus material. As such, any combination of Gan and Mechanic by a person of skill in the art would lead him or her to contemplate crosslinking the bioactive glass, polymer foam, or polymer foam coated with a sol gel bioactive material of Gan according to the methods of Mechanic. The combination would not lead the skilled artisan to the presently claimed invention. Applicant's arguments have been fully considered but they are not persuasive.

Firstly, the previous 35 U.S.C. 103(a) rejection of claims 82-88, 91-102 as being unpatentable over Mechanic in view of Gan et al. has been amended to being unpatentable over Gan et al. in view of Mechanic. No new art is being introduced in the instant action.

Regarding Applicants' assertions regarding the Gan et al. reference, Applicants are referred to column 8 of said reference. Gan et al. disclose intervertebral disc cells are isolated from tissue extracted from the nucleus pulposus of lumbar discs, sacral discs and cervical discs. The isolated cells are primarily nucleus pulposus cells which may be combined with a biodegradable substrate and implanted into the evacuated nucleus pulposus (col. 8 lines 50-58). However, Gan et al. further disclose that alternatively, the isolated nucleus pulposus cells can be cultured alone or seeded onto a biodegradable substrate and cultured together with said substrate for implantation (col. 8 lines 58-61). As previously noted, the term "tissue" as defined in the art, is an association of cells bound together by cell walls (plants) or extracellular matrix (animals) that performs a particular function. Therefore, one of ordinary skill would recognize that if cultured alone, the isolated nucleus pulposus cells of Gan et al. will expand into nucleus pulposus tissue, which can be used with a biodegradable substance for the manufacture of a disc implant. Further, as evidenced by Matsuzaki et al., since it is known that living nucleus pulposus cells synthesize collagen, it would be reasonable for one of ordinary skill to contemplate cross-linking the nucleus pulposus cells, and not to cross-link the biodegradable substance (as suggested by Applicants in their remarks) since cross-linking is recognized in the art as a method of stabilizing a protein matrix and preserving its structure and integrity.

Claims 89-90 are rejected under 35 U.S.C. 103(a) as being unpatentable over Gan et al. (US 5964807; previously cited) in view of Mechanic (US 5854397; previously cited) in view of Moore et al. (US 6350732; previously cited, IDS 11/03/06). The teachings of Gan et al. in view of Mechanic are outlined above. Neither Gan et al. nor Mechanic teach extracting lipids from a collagenous tissue sample.

Moore et al. teach a method for extracting lipids from collagenous tissue samples for the purpose of storing and preserving the tissue sample and the product of that method (col. 1 lines 29-35).

It would have been obvious to one of ordinary skill in the art to extract lipids from the nucleus pulposus matrix manufactured by the method of Gan et al. in view of Mechanic (claims 89-90). The motivation to do so is given by Moore et al., which teach that extracting lipids from collagenous tissue samples will allow the product to be better preserved and stored for longer periods of time. One of ordinary skill would recognize that nucleus pulposus cells and/or tissue that are better preserved would cause less complications when implanted in the body.

The reasons for maintaining the Gan et al. and Mechanic references are the same as noted above.

No claim is allowed.

THIS ACTION IS MADE FINAL. Applicant is reminded of the extension of time policy as set forth in 37 CFR 1.136(a).

A shortened statutory period for reply to this final action is set to expire THREE MONTHS from the mailing date of this action. In the event a first reply is filed within TWO MONTHS of the mailing date of this final action and the advisory action is not mailed until after the end of the THREE-MONTH shortened statutory period, then the shortened statutory period will expire on the date the advisory action is mailed, and any extension fee pursuant to 37 CFR 1.136(a) will be calculated from the mailing date of the advisory action. In no event, however, will the statutory period for reply expire later than SIX MONTHS from the mailing date of this final action.

Any inquiry concerning this communication or earlier communications from the examiner should be directed to Marsha M. Tsay whose telephone number is (571)272-2938. The examiner can normally be reached on M-F, 9:00am-5:00pm.

If attempts to reach the examiner by telephone are unsuccessful, the examiner's supervisor, Dr. Kathleen Kerr Bragdon can be reached on 571-272-0931. The fax phone number for the organization where this application or proceeding is assigned is 571-273-8300.

Information regarding the status of an application may be obtained from the Patent Application Information Retrieval (PAIR) system. Status information for published applications may be obtained from either Private PAIR or Public PAIR. Status information for unpublished applications is available through Private PAIR only. For more information about the PAIR system, see <http://pair-direct.uspto.gov>. Should you have questions on access to the Private PAIR system, contact the Electronic Business Center (EBC) at 866-217-9197 (toll-free). If you would

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like assistance from a USPTO Customer Service Representative or access to the automated information system, call 800-786-9199 (IN USA OR CANADA) or 571-272-1000.

/Maryam Monshipouri/

Primary Examiner, Art Unit 1656

March 20, 2008